

Vitamin D and periodontal disease

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Abstract: 1,25-Dihydroxyvitamin D₃ [1,25(OH)₂D₃; 1,25-dihydroxycholecalciferol or calcitriol] is the active form of vitamin D₃, a lipid-soluble vitamin that plays a role in calcium and bone metabolism. Recently, vitamin D₃ has been shown to function in cancer prevention, immunity and cardiovascular regulation. 1,25(OH)₂D₃ exhibits physiological and pharmacological effects by activating the vitamin D receptor (VDR), a transcription factor of the nuclear receptor superfamily. 1,25(OH)₂D₃ plays a role in maintaining oral health through its effects on bone and mineral metabolism and innate immunity, and several VDR gene polymorphisms have been reported to be associated with periodontal disease. VDR ligands should prove to be useful in the treatment and prevention of periodontal disease. (J Oral Sci 51, 11-20, 2009)

Keywords: vitamin D; vitamin D receptor; nuclear receptor; infection; innate immunity; periodontal disease.

Introduction

Vitamin D plays a role in various physiological processes, including bone and calcium metabolism, cellular growth and differentiation, immunity and cardiovascular function

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(1,2). Vitamin D is a secosteroid, in which the B ring of the canonical steroid structure is ruptured, and is synthesized from 7-dehydrocholesterol, an intermediate metabolite in cholesterol synthesis, or derived from dietary sources (3). Ultraviolet irradiation in sunlight-exposed skin induces a photochemical reaction of 7-dehydrocholesterol to produce the secosteroid vitamin D₃ (cholecalciferol) (Fig. 1A). Vitamin D₃ is hydroxylated at the 25-position by the hepatic vitamin D₃-hydroxylases, sterol 27-hydroxylase (CYP27A1) and vitamin D 25-hydroxylase (CYP2R1), to yield 25-hydroxyvitamin D₃ (25-hydroxycholecalciferol), the major form of vitamin D in the circulation (4). The 25-hydroxyvitamin D₃ is further hydroxylated in the 1 α -position by 25-hydroxyvitamin D 1 α -hydroxylase (CYP27B1). This reaction is tightly regulated and occurs exclusively in the kidney to yield the active metabolite, 1,25-dihydroxyvitamin D₃ [1,25(OH)₂D₃; 1,25-dihydroxycholecalciferol or calcitriol]. Dietary vitamin D₂ (ergocalciferol) and vitamin D₃ undergo the same activation process, involving 25-hydroxylation in the liver and subsequent 1 α -hydroxylation in the kidney, and are converted to the active metabolites, 1,25(OH)₂D₃ and 1,25(OH)₂D₂, respectively (3). These molecules bind to the vitamin D receptor (VDR), a nuclear receptor that is highly expressed in the target organs of calcium homeostasis, such as the intestine, bone, kidney and parathyroid glands (5). Recent epidemiological data and animal studies using VDR-null mice provide evidence for a role of vitamin D in preventing cancer, infection and cardiovascular disease as well as calcium and bone disorders (1,2,6,7) (Fig. 1B). This review focuses on the function of vitamin D in oral health.

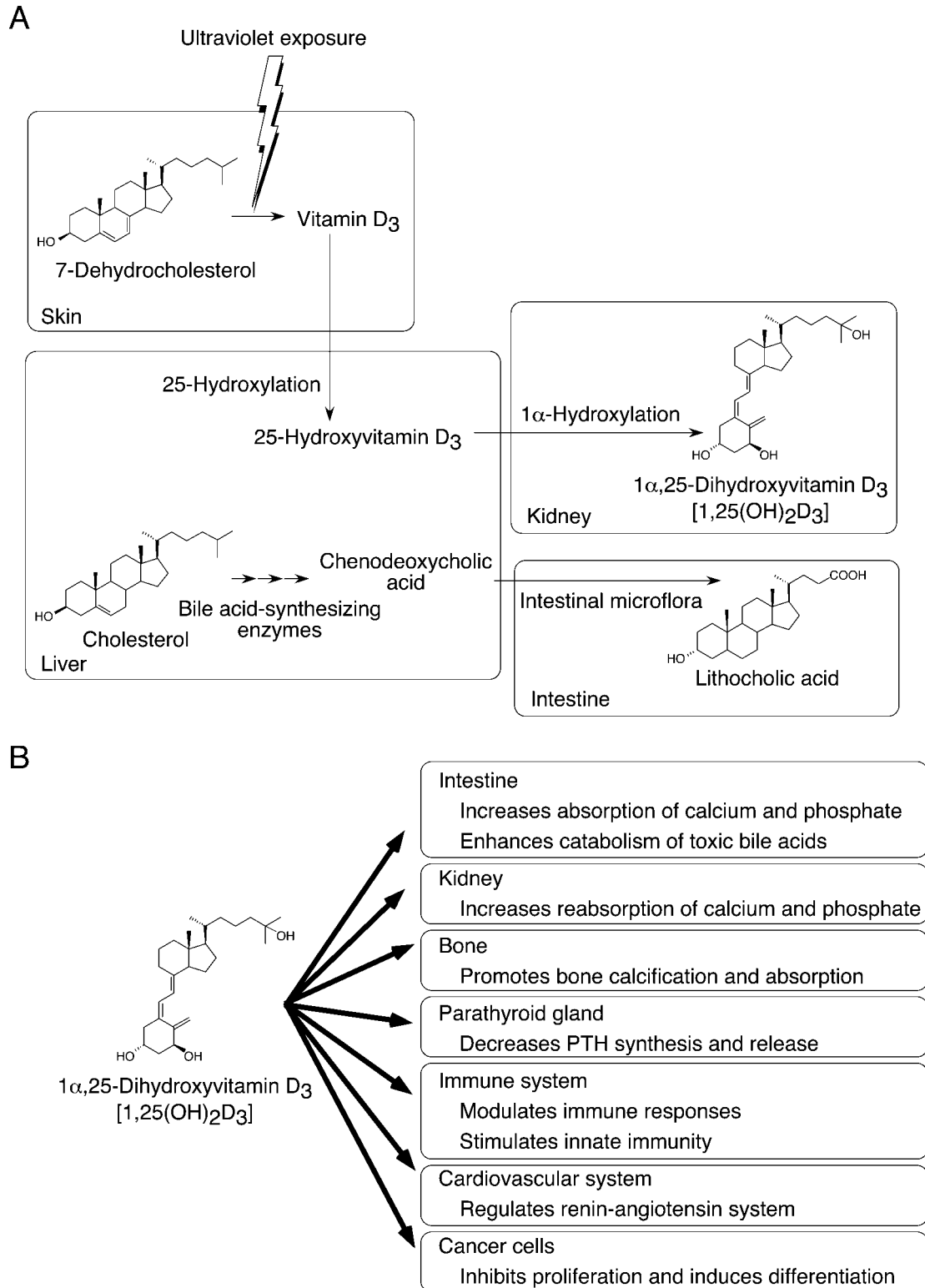


Fig. 1 Synthesis of natural VDR ligands and effects of 1,25(OH)₂D₃ on target cells. (A) 1,25(OH)₂D₃ is the active form of vitamin D₃ that activates VDR. 7-Dehydrocholesterol is converted to vitamin D₃ by ultraviolet-induced photochemical reaction, and then activated to 1,25(OH)₂D₃ by 25-hydroxylation in liver and 1α-hydroxylation in kidney. Cholesterol is metabolized to bile acids, such as chenodeoxycholic acid, in the liver. The primary bile acid chenodeoxycholic acid is converted to the secondary bile acid lithocholic acid, which is another natural VDR ligand. (B) 1,25(OH)₂D₃ exhibits physiological and pharmacological effects by activating VDR in target cells. The physiological role of lithocholic acid remains unknown.

Transactivation of VDR

VDR belongs to the nuclear receptor superfamily of transcription factor (8). Forty-eight human nuclear receptors have been identified and are classified into three groups on the basis of their ligand-binding characteristics. Steroid hormone receptors, which act as homodimers and mediate endocrine signals, are the first group and include the estrogen, progesterone, androgen, glucocorticoid and mineralocorticoid receptors. The second group are metabolic sensors and were initially identified as orphan receptors (9). Fatty acids, bile acids, oxysterols, and xenobiotics are ligands for this class of receptors. These metabolite-sensing receptors form heterodimers with retinoid X receptors (RXRs). The third group of orphan receptors have no known physiological ligands and may be regulated by ligand-independent mechanisms including phosphorylation. VDR responds to both an endocrine signal, $1,25(\text{OH})_2\text{D}_3$, and metabolites, such as lithocholic acid (Fig. 1A), indicating that VDR has dual functions as an endocrine receptor and a metabolic sensor (2). The organization of the human VDR protein, as in other nuclear receptors, has been divided into five regions (A-E) (Fig. 2A). The C region contains a DNA-binding domain with two zinc fingers and is the domain with strongest sequence homology among the member of the superfamily. The C-terminal ligand-binding domain (E region) forms a heterodimerization interface and contains a ligand-dependent transactivation domain called the activation function 2 (AF2). The N-terminal A/B region contains a ligand-independent transactivation domain called the AF1. AF1 domains play a role in tissue specific function of steroid hormone receptors, and the AF1 function of VDR may be limited because of its short A/B region (5).

VDR forms a heterodimer with RXR and is not permissive to RXR ligand activation (10). VDR is localized in both the cytosol and nucleus and accumulates in the nucleus in response to $1,25(\text{OH})_2\text{D}_3$ binding (11). The VDR-RXR heterodimer binds preferentially to a DNA response element that consists of a two hexanucleotide (AGGTCA or a related sequence) direct repeat separated by three nucleotides (DR3) (5) (Fig. 2B). The DR3 VDR-binding element has been identified in the regulatory regions of many target genes, including 25-hydroxyvitamin D 24-hydroxylase (CYP24A1), calbindin D_{9k} , cathelicidin antimicrobial peptide (CAMP) and transient receptor potential vanilloid type 6 (TRPV6). An everted repeat of the hexanucleotide motif separated by six nucleotides (ER6) is another VDR-binding element that regulates expression of the human CYP3A4 gene (12). Mutations in the zinc fingers of the VDR DNA-binding domain cause hereditary vitamin D-resistant rickets

(HVDRR) due to deficient target gene induction (5).

Nuclear receptors, including VDR, undergo a conformational change in the cofactor binding site and AF2 domain upon ligand binding, a structural rearrangement that results in the dynamic exchange of cofactor complexes (13). In the absence of ligand, corepressors bind to the AF2 surface, composed of portions of helix 3, loop 3-4, helices 4/5, and helix 11. Ligand binding alters the AF2 surface by repositioning helix 12 (Fig. 2B), reduces the affinity for corepressors, and increases the affinity for coactivator recruitment, allowing nuclear receptors to induce the transcription of specific genes. Cofactor complexes have been classified into three functional categories (14). Members of the first cofactor complex class regulate transcription directly via interactions with general transcription factors and RNA polymerase II. Members of the second cofactor complex class modify histone tails by acetylation or deacetylation. The third class of complexes is involved in ATP-dependent dynamic chromatin remodeling. Ligand-bound VDR is not only involved in transactivation but in some contexts can also mediate transrepression (15). Dynamic and coordinated interaction of cofactor complexes and VDR is required for efficient regulation of transcription.

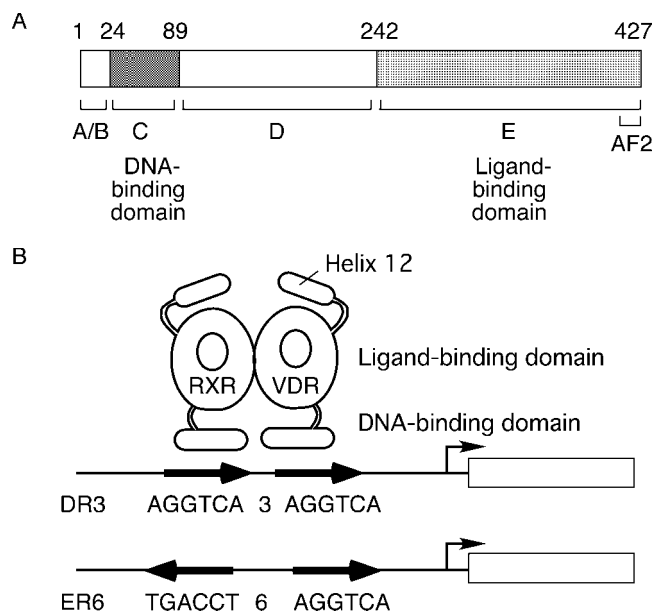


Fig. 2 VDR is a nuclear receptor mediating vitamin D signal. (A) Human VDR consists of 427 amino acids (GenBank accession no. NP_000367). (B) The VDR-RXR heterodimer binds to DR3 and ER6 elements in the promoter region of target genes. Helix 12 plays an important role in ligand-dependent activation by forming a cofactor interface.

Mineral and bone metabolism

$1,25(\text{OH})_2\text{D}_3$ plays an important role in maintaining calcium and phosphate levels in blood by stimulating intestinal absorption, bone resorption, and renal reabsorption (5) (Fig. 1B). Vitamin D deficiency causes insufficient absorption of dietary calcium and phosphate and results in increased secretion of parathyroid hormone to mobilize bone calcium stores, leading to rickets and osteomalacia. VDR mutations have been identified in HVDRR (16), and VDR-null mice have a similar phenotype to HVDRR patients, including rickets, hypocalcemia, hypophosphatemia, elevated serum $1,25(\text{OH})_2\text{D}_3$, and hyperparathyroidism (17). As in HVDRR patients, a high calcium diet prevents rickets and hyperparathyroidism in VDR-null mice (18). These findings indicate that the abnormal bone mineralization associated with vitamin D deficiency and HVDRR are secondary to impaired intestinal calcium absorption. Ligand-activated VDR induces the expression of genes involved in calcium metabolism, such as calbindin D_{9k} , TRPV6 and TRPV5 (19). Calbindin D_{9k} is an intracellular calcium transfer protein, and TRPV6 and TRPV5 are epithelial calcium channels. Although calbindin D_{9k} was considered to be an important mediator of vitamin D_3 signaling in renal and intestinal calcium absorption, mice lacking calbindin D_{9k} demonstrate that calbindin D_{9k} is not required for calcium homeostasis (20). TRPV6 is expressed in kidney and intestine, while TRPV5 expression is restricted to the kidney (19). Knockout mouse studies demonstrated that TRPV6 is necessary for intestinal calcium absorption and plays an important role in maintaining blood calcium levels (21). Mice lacking *Trpv5* have diminished renal calcium reabsorption, resulting in severe hypercalciuria (22). Through a compensatory increase in intestinal calcium absorption mediated by elevated serum vitamin D_3 levels and intestinal *Trpv6* expression, serum calcium levels are maintained. $1,25(\text{OH})_2\text{D}_3$ enhances dietary phosphate absorption by unknown mechanisms. Fibroblast growth factor 23 (FGF23) has been identified as a factor that reduces serum phosphate and $1,25(\text{OH})_2\text{D}_3$ levels by suppressing CYP27B1 expression and increasing CYP24A1 expression (23). Since VDR-null mice show normal skeletal development and their phenotype of rickets is rescued by normalization of serum calcium and phosphate levels, the effect of vitamin D deficiency on bone is mediated by dysregulated mineral homeostasis (24).

Activation of VDR by pharmacological doses of $1,25(\text{OH})_2\text{D}_3$ directly regulates osteoblasts by inducing the bone-remodeling proteins osteocalcin and osteopontin (5), and up-regulates the receptor activator of NF- κ B ligand (RANKL), a paracrine signal for osteoclastogenesis

(25). Transplantation of VDR-null bone to wild-type animals increases bone volume and density, indicating that VDR is involved in increased bone resorption or decreased bone formation (26). RANKL is a membrane-bound cytokine that binds to its receptor, RANK, which is expressed on osteoclast precursors, and activates osteoclast differentiation (27). VDR-mediated induction of osteoblast RANKL may account for enhanced bone resorption. Chondrocyte-specific VDR-ablated mice have reduced RANKL expression and delayed osteoclastogenesis (28). These mice also show reduced circulating levels of FGF23 and elevated serum phosphate levels. Since FGF23 is not expressed in chondrocytes, VDR induces an unknown chondrocyte-derived factor that up-regulates FGF23 expression in osteoblasts. Thus, VDR regulates bone homeostasis through actions in osteoblasts and chondrocytes as well as through mineral metabolism.

Osteoporosis is a common metabolic disease characterized by the loss of both the organic and mineral contents of bone, resulting in increased bone fragility and fracture. Osteoporosis and periodontal disease share several risk factors and bidirectional relationships between osteoporosis and periodontal disease have been proposed (29). Osteoporosis results in decreased bone mineral density throughout the body, including the maxilla and the mandible. The lowered density in the jawbones leads to increased alveolar porosity, an altered trabecular pattern and more rapid alveolar bone resorption following invasion by periodontal pathogens. Periodontal infection increases the systemic release of proinflammatory cytokines, which accelerate systemic bone resorption. Vitamin D deficiency is a risk factor for osteoporotic fractures (30), and treatment of osteoporotic women with $1,25(\text{OH})_2\text{D}_3$ increases bone mineral density and decreases the incidence of vertebral compression fractures (31). As discussed in the following sections, $1,25(\text{OH})_2\text{D}_3$ suppresses proinflammatory responses and enhances innate immunity. Therefore, VDR ligands should be clinically useful in the treatment of osteoporosis-associated periodontal disease.

Cancer and leukemia

$1,25(\text{OH})_2\text{D}_3$ has been demonstrated to inhibit the proliferation and induce differentiation of various types of malignant cells, including prostate, breast, colon, skin, and brain cancers, as well as myeloid leukemia cells *in vitro* (2). Epidemiological studies show an inverse relationship between mortality due to prostate, breast and colon cancers and sunlight exposure (6). Particularly in oral/pharyngeal cancer, low 25-hydroxyvitamin D_3 has been associated with increased cancer incidence, suggesting anticancer activity of $1,25(\text{OH})_2\text{D}_3$ (6,32). More than 20 years ago,

1,25(OH)₂D₃ was discovered to induce the differentiation of murine and human leukemia cells (33). Treatment with 1,25(OH)₂D₃ or 1 α -hydroxyvitamin D₃, which is rapidly metabolized to 1,25(OH)₂D₃, prolongs survival in leukemia mice (34). VDR ligands induce the expression of cyclin-dependent kinase inhibitors, p21^{CIP1/WAF1} and p27^{KIP1}, which may contribute to G1 cell cycle arrest of malignant cells (1). Although the anticancer mechanisms of 1,25(OH)₂D₃ remain to be elucidated, VDR ligands that retain efficient growth-inhibitory activity but have low calcemic activity should be promising anticancer drugs.

Immune disorders and infection

VDR is widely expressed in immune cells such as antigen-presenting cells, natural killer cells, T cells, and B cells, and 1,25(OH)₂D₃ has potent immunomodulatory effects (1). The immune effects of 1,25(OH)₂D₃ are principally mediated through actions on dendritic cells. Alloreactive T cell activation and dendritic cell maturation are inhibited by 1,25(OH)₂D₃. Hypertrophy due to increased mature dendritic cells in the lymph nodes of VDR deficient mice indicates that 1,25(OH)₂D₃ modulates antigen-specific immune responses *in vivo* (35). 1,25(OH)₂D₃ also has an effect on naïve CD4⁺ T cells to enhance Th2 cell development (36). Therapeutic effects of 1,25(OH)₂D₃ have been demonstrated in models of several immune diseases, including multiple sclerosis, rheumatoid arthritis, inflammatory bowel diseases, systemic lupus erythematosus, and transplant rejection (1,2).

Host-derived antimicrobial peptides participate in the innate immunity defense against mucosal infection (37). VDR activated by 1,25(OH)₂D₃ induces the expression of CAMP and β -defensin 4 and kills *Mycobacterium tuberculosis* in macrophages (38). Toll-like receptor activation by bacteria-derived lipopeptide up-regulates expression of VDR and CYP27B1 in macrophages, a mechanism that further enhances target gene induction. Reduced levels of 25-hydroxyvitamin D₃ in African Americans correlate with inefficient expression of CAMP mRNA and increased susceptibility to microbial infections. CAMP is widely expressed and secreted by keratinocytes and epidermal glands, and CAMP-deficient mice are susceptible to necrotic skin infection (39). β -Defensins exhibit antimicrobial activity against oral microbes including periodontitis-related bacteria such as *Actinobacillus actinomycetemcomitans*, *Porphyromonas gingivalis*, and *Fusobacterium nucleatum*, *Candida*, and papilloma virus (40). HVDRR patients suffer from frequent dental abscesses, providing further evidence that vitamin D plays a role in oral innate immunity (41).

VDR polymorphisms and periodontal disease

Along with the loss-of-function VDR mutations responsible for HVDRR (16), associations of several VDR restriction fragment length polymorphisms (RFLPs) with several diseases, including secondary hyperparathyroidism in renal failure, osteoporosis, cancer, nephrolithiasis, diabetes, and periodontal disease, have been reported (42-44). The RFLPs *BsmI*, *Tru9I*, *TaqI*, *EcoRV* and *ApaI* are located between exons 8 and 9 and may influence mRNA stability (45) (Fig. 3A). The RFLP *FokI* creates a start codon in exon 2, resulting in an alternative start site. An association between the *TaqI* RFLP (Fig. 3B) and periodontitis has been reported (46-48). An association between the less frequent t allele and localized early onset periodontitis (aggressive

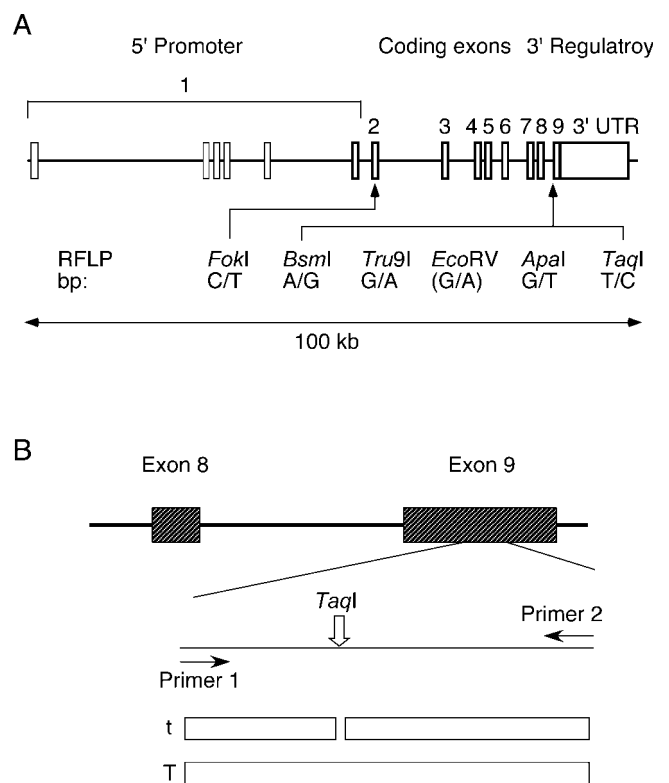


Fig. 3 Structure of the human VDR gene and VDR polymorphisms.

(A) RFLP sites *BsmI*, *Tru9I*, *EcoRV*, *ApaI* and *TaqI* are located between exon 8 and exon 9. *FokI* RFLP site is in exon 2. *FokI* and *TaqI* are in the coding sequence. (B) The *TaqI* RFLP is formed by a single base transition (T→C) at codon 352 in exon 9 of the VDR gene that creates a *TaqI* restriction site. The alleles that result from this change are designated "t" (*TaqI* site present) or "T" (*TaqI* site absent). Digestion of polymerase chain reaction fragments with *TaqI* results in different fragments for the t alleles and the T alleles (54).

Table 1 VDR polymorphisms and periodontitis

Authors, published year (reference)	Subjects	Country	Polymorphism	Association
Henning et al., 1999 (49)	69 EOP patients (including 20 LEOP) and 72 controls	UK	<i>TaqI</i>	LEOP: less frequent t allele
Yoshihara et al., 2001 (53)	42 GEOP patients, 52 AP patients and 55 healthy controls	Japan	<i>BsmI</i>	GEOP: VDR (B non-carrier) and FcgRIIIb (NA2 carrier) combination
Sun et al., 2002 (46)	24 AP patients and 37 EOP patients and 39 healthy controls	China	<i>TaqI</i>	EOP: Tt genotype and t allele
Tachi et al., 2003 (48)	74 CP patients and 94 healthy controls	Japan	<i>TaqI</i>	CP: TT genotype
de Brito Junior et al., 2004 (54)	69 CP patients and 44 healthy controls	Brazil	<i>TaqI, BsmI</i>	CP: TB haplotype and TB/tb heterozygous haplotype
Brett et al., 2005 (47)	51 AgP patients, 57 CP patients, and 100 healthy controls	UK	<i>TaqI</i>	CP: TT genotype
Park et al., 2006 (55)	93 GAgP patients and 143 healthy controls	Korea	<i>FokI</i>	GAgP: *C/*C genotype
Meng et al., 2007 (44)	90 AgP patients and 91 healthy controls	China	<i>TaqI</i>	AgP (female): Tt genotype

AP, adult periodontitis; AgP, aggressive periodontitis; CP, chronic periodontitis; EOP, early onset periodontitis; FcgRIIIb, immunoglobulin-Fcg receptor IIIb; GAgP, generalized aggressive periodontitis; GEOP, generalized early onset periodontitis; LEOP, localized early onset periodontitis

periodontitis) in Caucasian subjects was reported (49) (Table 1). The TT genotype and the T allele are associated with chronic periodontitis in Japanese and Caucasian subjects (47,48), while the tt genotype and t allele are associated with early onset periodontitis (aggressive periodontitis) in Chinese subjects (46). A strong association between Chinese female patients with aggressive periodontitis and the Tt genotype is suggested (44). While the tt genotype is associated with less occurrence of tuberculosis and chronic hepatitis B virus infection (50), the tt genotype and t allele are associated with decreases in bone mineral density and the incidence of osteoporosis (51,52). These findings suggest that the *TaqI* RFLP is associated with both immune function and bone metabolism. Ethnic differences and different mechanisms in pathogenesis between aggressive periodontitis and chronic periodontitis may influence the results of *TaqI* RFLP analysis. The *BsmI* RFLP in combination with other RFLPs are associated with early onset periodontitis (aggressive periodontitis) and chronic periodontitis (53,54). The *FokI* RFLP that results in the short VDR protein was demonstrated to increase a risk of generalized aggressive periodontitis in Korean (55). The *ApaI*, *BsmI*, and *FokI* RFLPs have been reported to confer elevated risk of severe

chronic periodontitis in Japanese men (56). These RFLPs are associated with bone and mineral diseases, and the *TaqI* and *FokI* RFLPs are associated with increased cancer risk, such as prostate and breast malignancy (42). Further studies are required to elucidate the functional relevance of VDR RFLPs and disease pathogenesis. An inverse association between serum 25-hydroxyvitamin D₃ concentrations and periodontal disease has been reported (57). These findings indicate that 1,25(OH)₂D₃ plays a role in prevention of periodontal disease and that hypomorphic VDR alleles and reduced levels of 1,25(OH)₂D₃ may be associated with periodontal disease.

Therapeutic application of VDR ligands

As discussed above, VDR ligands are promising drug candidates in the treatment of bone and mineral disorders, cancers and leukemia, autoimmune diseases and infection, including periodontal disease. Clinical studies have shown that vitamin D deficiency is associated with elevated risk of cardiovascular disease (7,58). Ligand-activated VDR suppresses renin expression and VDR-null mice develop cardiovascular disease, such as hypertension and cardiac hypertrophy, due to dysregulation of the renin-angiotensin system (59,60). VDR acts as a metabolic sensor for

secondary bile acids, such as lithocholic acid, and induces the expression of genes involved in the metabolism and excretion of toxic bile acids (2,61) (Fig. 1). These findings suggest that VDR-targeted therapies can be applied to cardiovascular disease and cholesterol/bile acid metabolism-related disorders. Insufficient clearance of periodontopathic bacteria and subsequent bone destruction are suggested to cause aggressive periodontitis (43). VDR ligands stimulate innate immunity by inducing antimicrobial peptides and have bone anabolic effects (1,38), suggesting that VDR ligands can be applied for prevention of aggressive periodontitis. A dysregulated release of proinflammatory cytokines by monocytes/macrophages and lymphocytes is considered to induce chronic periodontitis (43). Since $1,25(\text{OH})_2\text{D}_3$ has potent immunomodulatory effects, including inhibition of proinflammatory cytokine release (1,62), VDR ligands may be effective in the treatment of chronic periodontitis. Experimental and epidemiological studies suggest that VDR ligands are also useful in the prevention and treatment of oropharyngeal cancer (1,6). Currently, adverse effects, especially hypercalcemia, limit the clinical application of $1,25(\text{OH})_2\text{D}_3$ and its derivatives to bone and mineral disorders and psoriasis, a chronic skin disorder characterized by keratinocyte hyperproliferation and inflammatory infiltration of the epidermis and dermis (2). Combined dosing of $1,25(\text{OH})_2\text{D}_3$ with other drugs is one approach to overcome the adverse effects (63,64). The development of tissue-selective or function-selective VDR modulators with low calcemic activity provides another approach (65,66). Although topical application of VDR ligands, as in the treatment for psoriasis, may allow for the treatment of periodontal disease without inducing systemic adverse effects, further pharmacological and clinical studies are required.

Conclusion

In addition to its well-known activity in preventing rickets and osteomalacia, $1,25(\text{OH})_2\text{D}_3$ has been shown to have important anticancer, immune modulatory, and innate immune effects, through VDR activation. The $1,25(\text{OH})_2\text{D}_3$ -VDR system plays a role in oral homeostasis and its dysfunction may lead to periodontal disease. Vitamin D research should make important contributions to advancing oral medicine.

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References

1. Nagpal S, Na S, Rathnachalam R (2005) Noncalcemic actions of vitamin D receptor ligands. *Endocr Rev* 26, 662-687.
2. Makishima M, Yamada S (2005) Targeting the vitamin D receptor: advances in drug discovery. *Expert Opin Ther Pat* 15, 1133-1145.
3. Jones G, Strugnell SA, DeLuca HF (1998) Current understanding of the molecular actions of vitamin D. *Physiol Rev* 78, 1193-1231.
4. Cheng JB, Motola DL, Mangelsdorf DJ, Russell DW (2003) De-orphanization of cytochrome P450 2R1: a microsomal vitamin D 25-hydroxylase. *J Biol Chem* 278, 38084-38093.
5. Haussler MR, Whitfield GK, Haussler CA, Hsieh JC, Thompson PD, Selznick SH, Dominguez CE, Jurutka PW (1998) The nuclear vitamin D receptor: biological and molecular regulatory properties revealed. *J Bone Miner Res* 13, 325-349.
6. Giovannucci E, Liu Y, Rimm EB, Hollis BW, Fuchs CS, Stampfer MJ, Willett WC (2006) Prospective study of predictors of vitamin D status and cancer incidence and mortality in men. *J Natl Cancer Inst* 98, 451-459.
7. Wang TJ, Pencina MJ, Booth SL, Jacques PF, Ingelsson E, Lanier K, Benjamin EJ, D'Agostino RB, Wolf M, Vasan RS (2008) Vitamin D deficiency and risk of cardiovascular disease. *Circulation* 117, 503-511.
8. Makishima M (2005) Nuclear receptors as targets for drug development: regulation of cholesterol and bile acid metabolism by nuclear receptors. *J Pharmacol Sci* 97, 177-183.
9. Shulman AI, Mangelsdorf DJ (2005) Retinoid X receptor heterodimers in the metabolic syndrome. *N Engl J Med* 353, 604-615.
10. Shulman AI, Larson C, Mangelsdorf DJ, Ranganathan R (2004) Structural determinants of allosteric ligand activation in RXR heterodimers. *Cell* 116, 417-429.
11. Michigami T, Suga A, Yamazaki M, Shimizu C, Cai G, Okada S, Ozono K (1999) Identification of amino acid sequence in the hinge region of human vitamin D receptor that transfers a cytosolic protein to the nucleus. *J Biol Chem* 274, 33531-33538.
12. Thummel KE, Brimer C, Yasuda K, Thottassery J, Senn T, Lin Y, Ishizuka H, Kharasch E, Schuetz J, Schuetz E (2001) Transcriptional control of intestinal

- cytochrome P-4503A by $1\alpha,25$ -dihydroxy vitamin D₃. *Mol Pharmacol* 60, 1399-1406.
13. Rosenfeld MG, Lunyak VV, Glass CK (2006) Sensors and signals: a coactivator/corepressor/epigenetic code for integrating signal-dependent programs of transcriptional response. *Genes Dev* 20, 1405-1428.
 14. Kim M-S, Fujiki R, Kitagawa H, Kato S (2007) $1\alpha,25(\text{OH})_2\text{D}_3$ -induced DNA methylation suppresses the human CYP27B1 gene. *Mol Cell Endocrinol* 265-266, 168-173.
 15. Kato S, Fujiki R, Kim MS, Kitagawa H (2007) Ligand-induced transrepressive function of VDR requires a chromatin remodeling complex, WINAC. *J Steroid Biochem Mol Biol* 103, 372-380.
 16. Malloy PJ, Pike JW, Feldman D (1999) The vitamin D receptor and the syndrome of hereditary 1,25-dihydroxyvitamin D-resistant rickets. *Endocr Rev* 20, 156-188.
 17. Yoshizawa T, Handa Y, Uematsu Y, Takeda S, Sekine K, Yoshihara Y, Kawakami T, Arioka K, Sato H, Uchiyama Y, Masushige S, Fukamizu A, Matsumoto T, Kato S (1997) Mice lacking the vitamin D receptor exhibit impaired bone formation, uterine hypoplasia and growth retardation after weaning. *Nat Genet* 16, 391-396.
 18. Li YC, Amling M, Pirro AE, Priemel M, Meuse J, Baron R, Delling G, Demay MB (1998) Normalization of mineral ion homeostasis by dietary means prevents hyperparathyroidism, rickets, and osteomalacia, but not alopecia in vitamin D receptor-ablated mice. *Endocrinology* 139, 4391-4396.
 19. Nijenhuis T, Hoenderop JG, Bindels RJ (2005) TRPV5 and TRPV6 in Ca^{2+} (re)absorption: regulating Ca^{2+} entry at the gate. *Pflugers Arch* 451, 181-192.
 20. Kutuzova GD, Akhter S, Christakos S, Vanhooke J, Kimmel-Jehan C, DeLuca HF (2006) Calbindin D_{9k} knockout mice are indistinguishable from wild-type mice in phenotype and serum calcium level. *Proc Natl Acad Sci USA* 103, 12377-12381.
 21. Bianco SDC, Peng J-B, Takanaga H, Suzuki Y, Crescenzi A, Kos CH, Zhuang L, Freeman MR, Gouveia CHA, Wu J, Luo H, Mauro T, Brown EM, Hediger MA (2007) Marked disturbance of calcium homeostasis in mice with targeted disruption of the *Trpv6* calcium channel gene. *J Bone Miner Res* 22, 274-285.
 22. Hoenderop JGJ, van Leeuwen JPTM, van der Eerden BCJ, Kersten FFJ, van der Kemp AWCM, Merillat A-M, Waarsing JH, Rossier BC, Vallon V, Hummler E, Bindels RJM (2003) Renal Ca^{2+} wasting, hyperabsorption, and reduced bone thickness in mice lacking TRPV5. *J Clin Invest* 112, 1906-1914.
 23. Fukumoto S (2008) Physiological regulation and disorders of phosphate metabolism – pivotal role of fibroblast growth factor 23. *Intern Med* 47, 337-343.
 24. Amling M, Priemel M, Holzmann T, Chapin K, Rueger JM, Baron R, Demay MB (1999) Rescue of the skeletal phenotype of vitamin D receptor-ablated mice in the setting of normal mineral ion homeostasis: formal histomorphometric and biomechanical analyses. *Endocrinology* 140, 4982-4987.
 25. Kitazawa R, Kitazawa S (2002) Vitamin D₃ augments osteoclastogenesis via vitamin D-responsive element of mouse RANKL gene promoter. *Biochem Biophys Res Commun* 290, 650-655.
 26. Tanaka H, Seino Y (2004) Direct action of 1,25-dihydroxyvitamin D on bone: VDRKO bone shows excessive bone formation in normal mineral condition. *J Steroid Biochem Mol Biol* 89-90, 343-345.
 27. Yasuda H, Shima N, Nakagawa N, Yamaguchi K, Kinosaki M, Mochizuki S, Tomoyasu A, Yano K, Goto M, Murakami A, Tsuda E, Morinaga T, Higashio K, Udagawa N, Takahashi N, Suda T (1998) Osteoclast differentiation factor is a ligand for osteoprotegerin/osteoclastogenesis-inhibitory factor and is identical to TRANCE/RANKL. *Proc Natl Acad Sci USA* 95, 3597-3602.
 28. Masuyama R, Stockmans I, Torrekens S, Van Looveren R, Maes C, Carmeliet P, Bouillon R, Carmeliet G (2006) Vitamin D receptor in chondrocytes promotes osteoclastogenesis and regulates FGF23 production in osteoblasts. *J Clin Invest* 116, 3150-3159.
 29. Kuo L-C, Polson AM, Kang T (2008) Associations between periodontal diseases and systemic diseases: a review of the inter-relationships and interactions with diabetes, respiratory diseases, cardiovascular diseases and osteoporosis. *Public Health* 122, 417-433.
 30. van Schoor NM, Visser M, Pluijm SMF, Kuchuk N, Smit JH, Lips P (2008) Vitamin D deficiency as a risk factor for osteoporotic fractures. *Bone* 42, 260-266.
 31. Richey F, Ethgen O, Bruyere O, Reginster JY (2004) Efficacy of alfacalcidol and calcitriol in primary and corticosteroid-induced osteoporosis: a meta-analysis of their effects on bone mineral density and

- fracture rate. *Osteoporos Int* 15, 301-310.
32. Garland CF, Garland FC, Gorham ED (1999) Calcium and vitamin D. Their potential roles in colon and breast cancer prevention. *Ann N Y Acad Sci* 889, 107-119.
 33. Miyaoura C, Abe E, Kuribayashi T, Tanaka H, Konno K, Nishii Y, Suda T (1981) $1\alpha,25$ -Dihydroxyvitamin D₃ induces differentiation of human myeloid leukemia cells. *Biochem Biophys Res Commun* 102, 937-943.
 34. Honma Y, Hozumi M, Abe E, Konno K, Fukushima M, Hata S, Nishii Y, DeLuca HF, Suda T (1983) $1\alpha,25$ -Dihydroxyvitamin D₃ and 1α -hydroxyvitamin D₃ prolong survival time of mice inoculated with myeloid leukemia cells. *Proc Natl Acad Sci USA* 80, 201-204.
 35. Griffin MD, Lutz W, Phan VA, Bachman LA, McKean DJ, Kumar R (2001) Dendritic cell modulation by $1\alpha,25$ dihydroxyvitamin D₃ and its analogs: a vitamin D receptor-dependent pathway that promotes a persistent state of immaturity in vitro and in vivo. *Proc Natl Acad Sci USA* 98, 6800-6805.
 36. Boonstra A, Barrat FJ, Crain C, Heath VL, Savelkoul HFJ, O'Garra A (2001) $1\alpha,25$ -Dihydroxyvitamin D₃ has a direct effect on naive CD4⁺ T cells to enhance the development of Th2 cells. *J Immunol* 167, 4974-4980.
 37. Weinberg A, Krisanaprakornkit S, Dale BA (1998) Epithelial antimicrobial peptides: review and significance for oral applications. *Crit Rev Oral Biol Med* 9, 399-414.
 38. Liu PT, Stenger S, Li H, Wenzel L, Tan BH, Krutzik SR, Ochoa MT, Schaubert J, Wu K, Meinken C, Kamen DL, Wagner M, Bals R, Steinmeyer A, Zügel U, Gallo RL, Eisenberg D, Hewison M, Hollis BW, Adams JS, Bloom BR, Modlin RL (2006) Toll-like receptor triggering of a vitamin D-mediated human antimicrobial response. *Science* 311, 1770-1773.
 39. Nizet V, Ohtake T, Lauth X, Trowbridge J, Rudisill J, Dorschner RA, Pestonjamas V, Piraino J, Huttner K, Gallo RL (2001) Innate antimicrobial peptide protects the skin from invasive bacterial infection. *Nature* 414, 454-457.
 40. Abiko Y, Saitoh M, Nishimura M, Yamazaki M, Sawamura D, Kaku T (2007) Role of β -defensins in oral epithelial health and disease. *Med Mol Morphol* 40, 179-184.
 41. Seow WK (2003) Diagnosis and management of unusual dental abscesses in children. *Aust Dent J* 48, 156-168.
 42. Valdivielso JM, Fernandez E (2006) Vitamin D receptor polymorphisms and diseases. *Clin Chim Acta* 371, 1-12.
 43. Yoshie H, Kobayashi T, Tai H, Galicia JC (2007) The role of genetic polymorphisms in periodontitis. *Periodontol* 2000 43, 102-132.
 44. Meng H, Xu L, Li Q, Han J, Zhao Y (2007) Determinants of host susceptibility in aggressive periodontitis. *Periodontol* 2000 43, 133-159.
 45. Uitterlinden AG, Fang Y, van Meurs JBJ, Pols HAP, van Leeuwen JPTM (2004) Genetics and biology of vitamin D receptor polymorphisms. *Gene* 338, 143-156.
 46. Sun JL, Meng HX, Cao CF, Tachi Y, Shinohara M, Ueda M, Imai H, Ohura K (2002) Relationship between vitamin D receptor gene polymorphism and periodontitis. *J Periodontol* 37, 263-267.
 47. Brett PM, Zygianni P, Griffiths GS, Tomaz M, Parkar M, D'Aiuto F, Tonetti M (2005) Functional gene polymorphisms in aggressive and chronic periodontitis. *J Dent Res* 84, 1149-1153.
 48. Tachi Y, Shimpuku H, Nosaka Y, Kawamura T, Shinohara M, Ueda M, Imai H, Ohura K (2003) Vitamin D receptor gene polymorphism is associated with chronic periodontitis. *Life Sci* 73, 3313-3321.
 49. Hennig BJ, Parkhill JM, Chapple IL, Heasman PA, Taylor JJ (1999) Association of a vitamin D receptor gene polymorphism with localized early-onset periodontal diseases. *J Periodontol* 70, 1032-1038.
 50. Bellamy R, Ruwende C, Corrah T, McAdam KP, Thursz M, Whittle HC, Hill AV (1999) Tuberculosis and chronic hepatitis B virus infection in Africans and variation in the vitamin D receptor gene. *J Infect Dis* 179, 721-724.
 51. Spector TD, Keen RW, Arden NK, Morrison NA, Major PJ, Nguyen TV, Kelly PJ, Baker JR, Sambrook PN, Lanchbury JS, Eisman JA (1995) Influence of vitamin D receptor genotype on bone mineral density in postmenopausal women: a twin study in Britain. *BMJ* 310, 1357-1360.
 52. Zmuda JM, Cauley JA, Danielson ME, Wolf RL, Ferrell RE (1997) Vitamin D receptor gene polymorphisms, bone turnover, and rates of bone loss in older African-American women. *J Bone Miner Res* 12, 1446-1452.
 53. Yoshihara A, Sugita N, Yamamoto K, Kobayashi T, Miyazaki H, Yoshi H (2001) Analysis of vitamin D and Fc γ receptor polymorphisms in Japanese patients with generalized early-onset periodontitis. *J Dent Res* 80, 2051-2054.
 54. de Brito Junior RB, Scarel-Caminaga RM, Trevisatto

- PC, de Souza AP, Barros SP (2004) Polymorphisms in the vitamin D receptor gene are associated with periodontal disease. *J Periodontol* 75, 1090-1095.
55. Park KS, Nam JH, Choi J (2006) The short vitamin D receptor is associated with increased risk for generalized aggressive periodontitis. *J Clin Periodontol* 33, 524-528.
 56. Naito M, Miyaki K, Naito T, Zhang L, Hoshi K, Hara A, Masaki K, Tohyama S, Muramatsu M, Hamajima N, Nakayama T (2007) Association between vitamin D receptor gene haplotypes and chronic periodontitis among Japanese men. *Int J Med Sci* 4, 216-222.
 57. Dietrich T, Joshipura KJ, Dawson-Hughes B, Bischoff-Ferrari HA (2004) Association between serum concentrations of 25-hydroxyvitamin D₃ and periodontal disease in the US population. *Am J Clin Nutr* 80, 108-113.
 58. Giovannucci E, Liu Y, Hollis BW, Rimm EB (2008) 25-Hydroxyvitamin D and risk of myocardial infarction in men: a prospective study. *Arch Intern Med* 168, 1174-1180.
 59. Li YC, Kong J, Wei M, Chen Z-F, Liu SQ, Cao L-P (2002) 1,25-Dihydroxyvitamin D₃ is a negative endocrine regulator of the renin-angiotensin system. *J Clin Invest* 110, 229-238.
 60. Xiang W, Kong J, Chen S, Cao L-P, Qiao G, Zheng W, Liu W, Li X, Gardner DG, Li YC (2005) Cardiac hypertrophy in vitamin D receptor knockout mice: role of the systemic and cardiac renin-angiotensin systems. *Am J Physiol Endocrinol Metab* 288, E125-E132.
 61. Makishima M, Lu TT, Xie W, Whitfield GK, Domoto H, Evans RM, Haussler MR, Mangelsdorf DJ (2002) Vitamin D receptor as an intestinal bile acid sensor. *Science* 296, 1313-1316.
 62. Ogura M, Nishida S, Ishizawa M, Sakurai K, Shimizu M, Matsuo S, Amano S, Uno S, Makishima M (2009) Vitamin D₃ modulates the expression of bile acid regulatory genes and represses inflammation in bile duct-ligated mice. *J Pharmacol Exp Ther* 328, 564-570.
 63. Makishima M, Honma Y (1996) Ethacrynic acid and 1 α ,25-dihydroxyvitamin D₃ cooperatively inhibit proliferation and induce differentiation of human myeloid leukemia cells. *Leuk Res* 20, 781-789.
 64. Nakano H, Matsunawa M, Yasui A, Adachi R, Kawana K, Shimomura I, Makishima M (2005) Enhancement of ligand-dependent vitamin D receptor transactivation by the cardiotonic steroid bufalin. *Biochem Pharmacol* 70, 1479-1486.
 65. Ma Y, Khalifa B, Yee YK, Lu J, Memezawa A, Savkur RS, Yamamoto Y, Chintalacheruvu SR, Yamaoka K, Stayrook KR, Bramlett KS, Zeng QQ, Chandrasekhar S, Yu X-P, Linebarger JH, Iturria SJ, Burris TP, Kato S, Chin WW, Nagpal S (2006) Identification and characterization of noncalcemic, tissue-selective, nonsecosteroidal vitamin D receptor modulators. *J Clin Invest* 116, 892-904.
 66. Ishizawa M, Matsunawa M, Adachi R, Uno S, Ikeda K, Masuno H, Shimizu M, Iwasaki K, Yamada S, Makishima M (2008) Lithocholic acid derivatives act as selective vitamin D receptor modulators without inducing hypercalcemia. *J Lipid Res* 49, 763-772.